

The Effect of Dietary Constituents on Regulation of Epigenetic Changes in Cancer

S. Barghi (MSc)¹, M. Amiri (MSc)¹, H. Hajipour (PhD)², S. Namaki (PhD)^{*3}

1. Department of Medical Laboratory Sciences, Faculty of Paramedical Sciences, Shahid Beheshti University of Medical Science, Tehran, I.R.Iran

2. Department of Reproductive Biology, Faculty of Advanced Medical Sciences, Tabriz University of Medical Sciences, Tabriz, I.R.Iran

3. Department of Immunology, Faculty of Medical Sciences, Shahid Beheshti University of Medical Sciences, Tehran, I.R.Iran

J Babol Univ Med Sci; 19(7); Jul 2017; PP: 63-71

Received: Nov 10th 2016, Revised: Jan 25th 2017, Accepted: May 10th 2017.

ABSTRACT

BACKGROUND AND OBJECTIVE: The term “epigenetic” refers to all non-heritable and reversible changes in the expression of a gene that does not cause a change in the DNA sequence. The most important epigenetic mechanisms associated with gene expression include DNA methylation, histone modifications, and suppression of gene expression with RNA. Considering the reversibility of epigenetic changes, it seems that this feature can be influenced by dietary constituents and thus, we can prevent the spread of certain cancers by controlling the diet. The purpose of this study is to investigate the effects of food on the prevention of common cancers and the mechanisms involved in cellular activities based on recent studies and the compilation of their results.

METHODS: In this review article, we searched Pubmed and Elsevier databases using certain keywords such as “epigenetics”, “cancer” and “nutrition” and articles related to the effects of epigenetics on cancer and dietary constituents were evaluated.

FINDINGS: Of 439 studies found in the search engines between 1997 and 2016, 64 articles were selected and their results indicated that many of the active components in the diet will inhibit the incidence of cancer through DNA methylation mechanisms, histone modifications, and miRNA.

CONCLUSION: The anticancer effect of the active compounds in the diet on specific epigenetic changes can be used as a special and unidentified mechanism for preventing cancer.

KEY WORDS: *Epigenetic, Cancer, Nutrition.*

Please cite this article as follows:

Barghi S, Amiri M, Hajipour H, Namaki S. The Effect of Dietary Constituents on Regulation of Epigenetic Changes in Cancer. J Babol Univ Med Sci. 2017;19(7):63-71.

* Corresponding author: S. Namaki (PhD)

Address: Department of Immunology, Faculty of Medical Sciences, Shahid Beheshti University of Medical Sciences, Tehran, I.R. Iran

Tel: +98 21 22718531

E-mail: saeednamaki@yahoo.com

Introduction

Epigenetic changes are inherited changes in gene expression that do not change the DNA sequence (1). There is a lot of evidence indicating that there is a relationship between epigenetic disorders and various diseases, including cancer, multiple sclerosis (2), diabetes (3), and obesity. Recent studies show the effects of dietary compounds on the cancer process. The isothiocyanate in the family of cabbage (cauliflower, broccoli), diallyl sulfide (organosulfur compound in garlic), isoflavone, phytosterol, theophylline, folate, selenium, vitamin E, flavonoids and dietary fiber may reduce the risk of cancer (4, 5). The main mechanisms of epigenetic control in mammals include DNA methylation, histone modifications, and RNA interferences (6, 7).

The key to epigenetic changes in mammals is the addition of a 5-methylcytosine group in two nucleotide sequences of CPG (1, 8). The CPG sequences are CG-rich regions known as CPG islands and are associated with transcriptional initiation regions (6, 8). Hypermethylation of these regions can lead to silencing of transcription of tumor suppressor genes and their inactivation in various cancers (1).

The addition of covalent methyl group is catalyzed by DNA methyltransferase family (DNMTs), which utilizes S-Adenosyl methionine (SAM) as a methyl group provider (8). DNMT1 is initially involved in the storage of DNA methylation after replication, and DNMT3A and DNMT3B interact with the transcriptional machine and mediate methylation. Several studies have shown that DNMTs can be expressed in a number of cancers (8). The purpose of this study is to investigate the effects of food on the prevention of common cancers and the mechanisms involved in cellular activities based on recent studies and the compilation of their results.

Methods

In this review article, the effect of food on the cancer formation process using the key words "Epigenetic", "Cancer" and "Nutrition", the scientific articles indexed in Pubmed and Elsevier databases between 1997 and 2016 were reviewed and the articles were carefully evaluated.

Results

439 articles were extracted in relation to the keywords, among which 64 articles were selected and reviewed.

Articles that examined the effects of other factors such as age or the presence of environmental factors such as UV were excluded. According to these studies, active food compound cause epigenetic changes with mechanisms such as DNA methylation in progesterone CPGs, histone modifications, and miRNA expression.

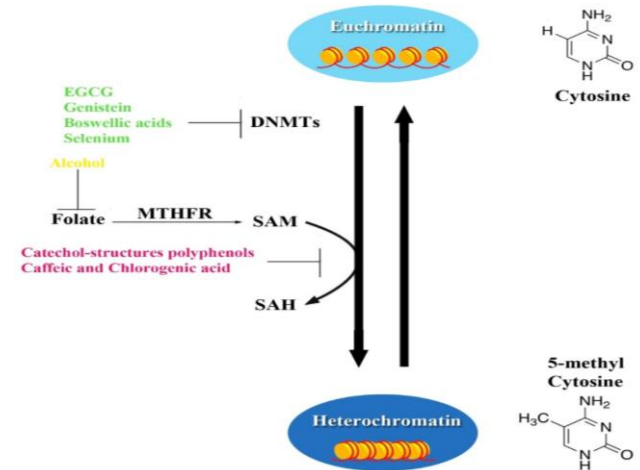


Figure 1. DNA methylation in the cytosine of the CPG island promoters leads to silencing of transcription of tumor suppressor genes and their malignant changes

Dietary compounds, DNA methylation and hereditary epigenetic changes: A number of epidemiological studies associate adverse environmental and nutritional conditions in the early stages of growth and embryonic development, and then at puberty, with the risk of developing certain diseases in adulthood. Although the underlying mechanism of this relationship is still unknown, evidence suggests interference with epigenetic disorders (8–11).

Dietary compounds and DNA methylation in cancer: Compounds such as folate, tea polyphenols, soy isoflavones and catechol polyphenols have anti-cancer properties with DNA methylation mechanisms (12, 13) (Fig 1). Folate is involved in the metabolism of single-carbon units, DNA synthesis and DNA methylation (14). Folate deficiency causes cancer through DNA damage (inappropriate uracil pairing) (15, 16), ectopic methylation, such as promoter methylation (17, 18), and DNMT1 inhibition (19). Enriching a diet with folic acid or natural folate reduces the risk of colorectal cancer (20, 21). Additionally, its high doses (20 mg folate/kg) significantly decrease the polyps of the intestine in *Apc^{Min/+}* mice after 3 months. However, after 6 months, supplementation with folate has an opposite effect on the number of polyps in the intestine (22). In a study, folic acid deficiency in mothers was

associated with DNA ectopic methylation, which leads to neural tube defects. Low levels of folate in the serum of mothers are associated with DNA hypomethylation in the brain and DNA hypermethylation in the skin and fetal heart that is associated with neural tube defects (23).

EGCG is the main polyphenol in green tea with antioxidant properties that can inhibit tumor metastasis and angiogenesis (24). Epidemiological studies have shown that using green tea reduces the risk of hepatocellular carcinoma (25). EGCG directly or indirectly inhibits DNMT (26). Treatment of human cancer cells, esophageal cancer cells KYSE510, HT-29 colon cancer, and PC3-induced prostate cancer with EGCG result in the return of hypermethylation of p16, RAR β , and MGMT genes (27).

Treatment of Caco-2 cells with EGCG inhibits cellular growth and inhibits the promoter's methylation of anti-tumor genes p16 and P15 (28). Treatment of breast cancer cell line MCF-7 and promyelocytic leukemia cells with EGCG have the potential to reduce cell proliferation and induction of apoptosis (29). In fact, EGCG is effective in both antioxidant and epigenetic changes to various cancer cells. However, the potential harmful effects of high consumption of green tea (DNA hypomethylation and oncogene activation and genomic instability) should be taken into account.

Genistein (soy isoflavone) has the effects of cancer prevention through epigenetic mechanisms (30). Genistein causes the reversal of DNA methylation and re-activation of RAR β and MGMT genes in KYSE510 cells (31), and at high doses, it inhibits the DNA of methyl transferases in LNCap and PC3 cell lines in prostate cancer (32).

These findings indicate that genistein activates tumor suppressor genes that have been silenced. Resveratrol is a natural phytoalexin compound that the ability to inhibit proliferation of cells, which increases the methylation of P16 and reduces the methylation of P15 in Caco2 cells (28).

Curcumin (Curcuma longa rhizome flavonoid) also has anticancer activity. Curcumin and genistein cause the reversal of the RAR β 2 gene hypermethylation in the cervical cancer cell line SiHa and HeLa, as well as progressive demethylation (33). Quercetin is a flavonoid with antioxidant and anti-proliferative activity, which is a natural inhibitor of catechol-O-methyl transferase (COMT). Quercetin induces cell cycle stoppage and apoptosis of the hamster's oral

tumor, and the effect is related to the control of DNMT1 (33). Quercetin also increases the bioavailability of green tea polyphenols in the investigation of the A549 and O-786 cell lines, as well as in mice with immunodeficiency (34).

In addition, quercetin boosts anti-proliferative activity of EGCG by increasing intracellular EGCG concentration and reducing methylation in prostate cancer cells (35). Selenium is an essential ingredient that has potential for preventing cancer due to antioxidant and pro-apoptotic effects (31, 36). Treatment of Caco2 cells with selenite induces total hypomethylation and promoter methylation of the P53 gene (36).

Selenium plays an anticancer role in human colon cancer through DNMT inhibition (37). However, selenium and vitamin E inhibitors did not provide evidence to prove that selenium prevents prostate, lung, or colorectal cancers (38).

Changes after histone translation: Histones have an active function in regulating chromatin structure and gene expression. Histone tails may change by acetylation, Methylation, Phosphorylation, Poly ADP-Ribosylation, sumoylation, or ubiquitination (6, 39). DNA methylation and histone modification are not independent events.

The methylation of cytosine in CPG islands is associated with the attachment between binding proteins and methyl cytosine, followed by catalytic enzymes of histone modifications (1, 6). Acetylation in histone lysine amino group neutralizes the positive lysine load by histone acetyltransferases (HATs) and releases the histone tail of the negatively charged DNA. These changes lead to access to transcription factors for the expression of genes in that area (6). Histone deacetylases leads to chromatin congestion and transcription inhibition by histone deacetylases (HDACs). In the deacetylated state, the amine lysine group has a positive charge and allows the tail of the histone to interact strongly with the DNA strand that has a negative charge (40, 41).

Methylation of the roots of lysine and arginine in H3 and H4 histones can have suppressive and transcriptional effects depending on the type of amino acid and its position. Histone methylation without changing the histone load changes its chemical properties and thus affects the tendency for regulatory proteins. Histone methylation is catalyzed by histone methyltransferases, while the omission of methyl groups is catalyzed by histone demethylase (42)

(Fig2). H1 phosphorylation weakens its binding to DNA and gives free access to transcription factors and gene expression. Histone phosphorylation is observed in a number of cancers, such as breast, prostate and colorectal cancer (39).

Dietary compounds and histone modifications in cancer: Folate can affect the methylation of histones in cancer. A diet that is low in methyl donors leads to changes in H4-K20 methylation and H3-K9 acetylation, which is usually observed in liver cancer (43). EGCG induces changes in human A431 melanoma cells. EGCG reduces the deacetylation activity of histone and increases lysine acetylation in histones 4 and 3 and reduces lysine 9 histone 3 methylation (44).

In the MCF-7 cells, genistein reduces the histone 3 acetylation and induces growth response to mitogens and histone deacetylase (HDAC) inhibitors (45). In the MCF-7 breast cancer cell line, resveratrol inhibits the dioxin-induced histone modifications in the BRCA-1 gene and suppresses the expression of BRCA1 protein, decreasing the DNA fragmentation associated with dioxin (46).

Treatment of brain cancer cells with curcumin causes hypomethylation of H3 and H4 histones (47). Conversely, in prostate cancer cells, curcumin induces acetylation of H3 and H4 and also causes apoptosis through Bcl2 and P53 families (48).

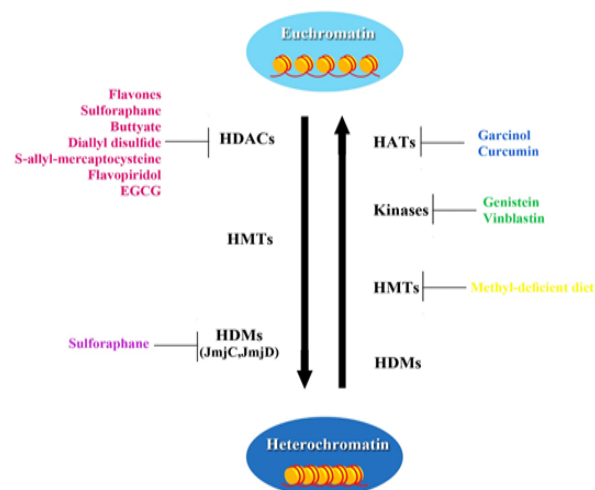


Figure 2. Histone modifications determine the interaction of histones with DNA and the interaction of non-histone proteins with chromatin

RNA-related silence by microRNAs in cancer:

Micro-RNAs or miRNAs play a role in the post-transcriptional configuration by binding to the non-translatable region of the target 3' (3'UTR) mRNA (49,

50). MiRNA acts in two ways: full pairing with complementary cells that results in the destruction of the target mRNA, and partial pairing that results in inhibiting the translation of the target mRNA (49, 50) (Fig 3). In addition, they also play a role in transcriptional regulation, which can bind to complementary sequences in the genome and induce gene silencing through the administration of suppressor proteins and induction of symptoms of chromatin suppression (51, 52). Changes in the expression of miRNAs are a common occurrence in cancer. Some of them act as suppressor tumors of hypothetical genes, and some as oncogenes (49, 50).

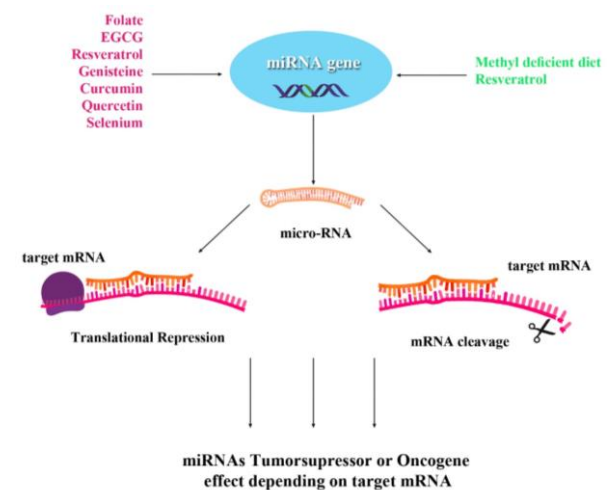


Figure 3. MiRNAs are small and noncoding RNAs that contribute to post-transcription regulation through binding to the target mRNA

Dietary components and miRNA changes in cancer:

Nutrition, lifestyle and genetic factors also affect the risk of cancer through miRNAs (Fig. 3). Fat-rich diets in mothers before fertilization, during pregnancy and breastfeeding induces long-term changes in IGF2 expression (11). Folate deficiency leads to a general increase in the expression of miRNA in lymphocyte (53). The hepatocarcinogenesis of methyl deficiency in mice results in reduced expression of miRNA gene (54, 55), including miRNA-34a and miRNA-127, which are involved in apoptosis and cell proliferation, respectively (56).

In addition, folic acid supplement inhibits the expression of miRNA-10a induced by alcohol use (57). EGCG modifies the expression of a number of miRNAs in human hepatocyte carcinoma, including miRNA-16, which targets the anti-apoptotic protein Bcl-2 (58). Treatment with uveal melanoma cells with genistein decreases cell proliferation and increases the

expression of miRNA-27a (59). In BALB / C nu / nu mice, genistein significantly inhibits the growth of the human uveal melanoma xenografts (59). The treatment of the colon cancer cell line SW480 with resveratrol reduces the level of several oncogene miRNAs targeting PTEN and PDCD4 (60). Simultaneously, this treatment increases the amount of miR-663 that targets the TGF β growth factor transcript (61).

Treatment of BxPC-3 human pancreatic cancer cells with curcumin increases the expression of miRNA22 and decreases expression of miRNA-199a (62). Quercetin induces miR-146a, a negative regulator of pro-inflammatory NF-kB activity in HT-29 colon cancer cells (63). The treatment of LNCaP prostate cancer cells with selenite-induced p53 induces apoptosis and expresses miR-34 that targets the P53 transcript (64).

Discussion

This study shows that active compounds are effective on DNA methylation, histone modifications, and miRNA expression in cancer. However, it is unlikely that the protective effects would be caused by only one dietary component. Therefore, identification of relevant compounds and metabolites is required. Another key issue is the appropriate concentrations of

herbal compounds for inducing optimal epigenetic modifications (6).

Epigenetic changes in tissue are important for cellular differentiation. Thus, active food components may cause different epigenetic changes in different tissues and even different types of cells in a tissue. In addition, epigenetic changes induced by food components can be temporary. Therefore, it is important to recognize the specific effects of the cell and tissue of an active dietary compound and related kinetics. The development of relevant animal tissue culture models is necessary to study the effects of diet and the environment on epigenetic changes to clarify their relationship and their interaction potential.

The complex interactions between environmental, genetic and epigenetic factors during cancer development have not been thoroughly defined. However, the definition of bioactive food components is necessary to provide safe dietary advice and to determine the dose required for preventative effects. Therefore, further studies are needed to identify epigenetic changes and cellular features and time patterns. Nevertheless, despite so many unresolved questions, there is a promising future for dietary recommendations on cancer prevention and the provision of natural-based treatment programs for cancer treatment.

References

- 1.Herman JG, Baylin SB. Gene silencing in cancer in association with promoter hypermethylation. *N Engl J Med*. 2003;349(21):2042-54.
- 2.Mirshafiey A, Aghily B, Namaki S, Razavi A, Ghazavi A, Ekhtiari P, et al. Therapeutic approach by Aloe vera in experimental model of multiple sclerosis. *Immunopharmacol Immunotoxicol*. 2010;32(3):410-5.
- 3.Safari-Alighiarloo N, Taghizadeh M, Tabatabaei SM, Shahsavari S, Namaki S, Khodakarim S, et al. Identification of new key genes for type 1 diabetes through construction and analysis of protein–protein interaction networks based on blood and pancreatic islet transcriptomes. *J Diabet*. 2017;9(8):764-77.
- 4.Amiri M, Kazerouni F, Namaki S, Tamijani HD, Rahimipour H, Boroumand N, et al. Cytotoxic Effects of the Ethanol Bane Skin Extract in Human Prostate Cancer Pc3 Cells. *Iran J Cancer Prevention*. 2016;9(2): 4755.
- 5.Saboury AA, Bagheri S, Ataie G, Amanlou M, Moosavi-Movahedi AA, Hakimelahi GH, et al. Binding properties of adenosine deaminase interacted with theophylline. *Chem Pharma bulletin*. 2004;52(10):1179-82.
- 6.Laird PW. Cancer epigenetics. *Hum Molecul Gen*. 2005;14(1):65-76.
- 7.Donovan MG, Selmin OI, Doetschman TC, Romagnolo DF. Mediterranean Diet, Inflammatory Bowel Diseases, and Colon Cancer. *Mediterranean Diet: Springer*; 2016. p. 181-201.
- 8.Tobi EW, Lumey L, Talens RP, Kremer D, Putter H, Stein AD, et al. DNA methylation differences after exposure to prenatal famine are common and timing-and sex-specific. *Hum Mol Genet*. 2009;18(21):4046-53.
- 9.Hughes LA, van den Brandt PA, De Bruïne AP, Wouters KA, Hulsmans S, Spiertz A, et al. Early life exposure to famine and colorectal cancer risk: a role for epigenetic mechanisms. *PloS One*. 2009;4(11):7951.
- 10.Tobi EW. Persistent epigenetic differences associated with prenatal exposure to famine in humans. *Dutch Hung Win*. 2008;105(44):55.
- 11.Zhang J, Zhang F, Didelot X, Bruce KD, Cagampang FR, Vavish M, et al. Maternal high fat diet during pregnancy and lactation alters hepatic expression of insulin like growth factor-2 and key microRNAs in the adult offspring. *BMC Gen*. 2009;10(1):1.
- 12.Yoon J-H, Baek SJ. Molecular targets of dietary polyphenols with anti-inflammatory properties. *Yonsei Med J*. 2005;46(5):585-96.
- 13.Yang CS, Landau JM, Huang M-T, Newmark HL. Inhibition of carcinogenesis by dietary polyphenolic compounds. *Ann Rev Nut*. 2001;21(1):381-406.
- 14.Kim Y-I. Folate and carcinogenesis: evidence, mechanisms, and implications. *J Nut Biochem*. 1999;10(2):66-88.
- 15.van den Donk M, Pellis L, Crott JW, van Engeland M, Friederich P, Nagengast FM, et al. Folic acid and vitamin B-12 supplementation does not favorably influence uracil incorporation and promoter methylation in rectal mucosa DNA of subjects with previous colorectal adenomas. *J Nut*. 2007;137(9):2114-20.
- 16.Duthie SJ, Narayanan S, Blum S, Pirie L, Brand GM. Folate deficiency in vitro induces uracil misincorporation and DNA hypomethylation and inhibits DNA excision repair in immortalized normal human colon epithelial cells. *Nut Cancer*. 2000;37(2):245-51.
- 17.Kim Y-I. Folate and DNA methylation: a mechanistic link between folate deficiency and colorectal cancer?. *Can Epidemiol Biomark Prevent*. 2004;13(4):511-9.
- 18.van Engeland M, Weijenberg MP, Roemen GM, Brink M, de Bruïne AP, Goldbohm RA, et al. Effects of dietary folate and alcohol intake on promoter methylation in sporadic colorectal cancer: the Netherlands cohort study on diet and cancer. *Cancer Res*. 2003;63(12):3133-7.
- 19.Trasler J, Deng L, Melnyk S, Pogribny I, Hiou-Tim F, Sibani S, et al. Impact of Dnmt1 deficiency, with and without low folate diets, on tumor numbers and DNA methylation in Min mice. *Carcinogenesis*. 2003;24(1):39-45.
- 20.Giovannucci E, Stampfer MJ, Colditz GA, Hunter DJ, Fuchs C, Rosner BA, et al. Multivitamin use, folate, and colon cancer in women in the Nurses' Health Study. *Ann Inter Med*. 1998;129(7):517-24.

21. Sanjoaquin MA, Allen N, Couto E, Roddam AW, Key TJ. Folate intake and colorectal cancer risk: a meta-analytical approach. *Inter J Cancer*. 2005;113(5):825-8.
22. Song J, Medline A, Mason JB, Gallinger S, Kim Y-I. Effects of dietary folate on intestinal tumorigenesis in the *apcMin* mouse. *Cancer Res*. 2000;60(19):5434-40.
23. Chang H, Zhang T, Zhang Z, Bao R, Fu C, Wang Z, et al. Tissue-specific distribution of aberrant DNA methylation associated with maternal low-folate status in human neural tube defects. *J Nut Biochem*. 2011;22(12):1172-7.
24. Hajipour H, Hamishehkar H, Raeisi S, Barghi S, Hasani A. Epigallocatechin-3-Gallate Induces Apoptosis through Up-regulation of Bax and Down-regulation of Bcl-2 in Prostate Cancer Cell Line. *Inter J Med Lab*. 2016;3(4):262-9.
25. Khan N, Mukhtar H. Cancer and metastasis: prevention and treatment by green tea. *Cancer Metastasis Rev*. 2010;29(3):435-45.
26. Lu H, Meng X, Li C, Sang S, Patten C, Sheng S, et al. Glucuronides of tea catechins: enzymology of biosynthesis and biological activities. *Drug Metabolism and Disposition*. 2003;31(4):452-61.
27. Fang MZ, Wang Y, Ai N, Hou Z, Sun Y, Lu H, et al. Tea polyphenol (-)-epigallocatechin-3-gallate inhibits DNA methyltransferase and reactivates methylation-silenced genes in cancer cell lines. *Cancer Res*. 2003;63(22):7563-70.
28. Berner C, Aumüller E, Gnauck A, Nestelberger M, Just A, Haslberger AG. Epigenetic control of estrogen receptor expression and tumor suppressor genes is modulated by bioactive food compounds. *Ann Nut Metabol*. 2010;57(3-4):183-9.
29. Berletch JB, Liu C, Love WK, Andrews LG, Katiyar SK, Tollefsbol TO. Epigenetic and genetic mechanisms contribute to telomerase inhibition by EGCG. *J Cell Biochem*. 2008;103(2):509-19.
30. Li Y, Tollefsbol TO. Impact on DNA methylation in cancer prevention and therapy by bioactive dietary components. *Curr Med Chem*. 2010;17(20):2141.
31. Barrera LN, Cassidy A, Johnson IT, Bao Y, Belshaw NJ. Epigenetic and antioxidant effects of dietary isothiocyanates and selenium: potential implications for cancer chemoprevention. *Proc Nut Soc*. 2012;71(02):237-45.
32. Fang MZ, Chen D, Sun Y, Jin Z, Christman JK, Yang CS. Reversal of hypermethylation and reactivation of p16INK4a, RAR β , and MGMT genes by genistein and other isoflavones from soy. *Clin Cancer Res*. 2005;11(19):7033-41.
33. Priyadarsini RV, Vinothini G, Murugan RS, Manikandan P, Nagini S. The flavonoid quercetin modulates the hallmark capabilities of hamster buccal pouch tumors. *Nutrition Cancer*. 2011;63(2):218-26.
34. Wang P, Heber D, Henning SM. Quercetin increased bioavailability and decreased methylation of green tea polyphenols in vitro and in vivo. *Food Fun*. 2012;3(6):635-42.
35. Wang P, Heber D, Henning SM. Quercetin increased the antiproliferative activity of green tea polyphenol (-)-epigallocatechin gallate in prostate cancer cells. *Nut Cancer*. 2012;64(4):580-7.
36. Davis CD, Uthus EO, Finley JW. Dietary selenium and arsenic affect DNA methylation in vitro in Caco-2 cells and in vivo in rat liver and colon. *J Nut*. 2000;130(12):2903-9.
37. Fiala ES, Staretz ME, Pandya GA, El-Bayoumy K, Hamilton SR. Inhibition of DNA cytosine methyltransferase by chemopreventive selenium compounds, determined by an improved assay for DNA cytosine methyltransferase and DNA cytosine methylation. *Carcinogenesis*. 1998;19(4):597-604.
38. Lippman SM, Klein EA, Goodman PJ, Lucia MS, Thompson IM, Ford LG, et al. Effect of selenium and vitamin E on risk of prostate cancer and other cancers: the Selenium and Vitamin E Cancer Prevention Trial (SELECT). *JAMA*. 2009;301(1):39-51.
39. Jenuwein T, Allis CD. Translating the histone code. *Science*. 2001;293(5532):1074-80.
40. Margueron R, Trojer P, Reinberg D. The key to development: interpreting the histone code?. *Curr Opin Genet Dev*. 2005;15(2):163-76.

41. Mählkecht U, Hoelzer D. Histone acetylation modifiers in the pathogenesis of malignant disease. *Mol Med.* 2000;6(8):623.
42. Miremadi A, Oestergaard MZ, Pharoah PD, Caldas C. Cancer genetics of epigenetic genes. *Hum Mol Gen.* 2007;16(1):28-49.
43. Pogribny IP, Tryndyak VP, Muskhelishvili L, Rusyn I, Ross SA. Methyl deficiency, alterations in global histone modifications, and carcinogenesis. *J Nut.* 2007;137(1):216S-21S.
44. Andakumar V, Vaid M, Katiyar SK. (-)-Epigallocatechin-3-gallate reactivates silenced tumor suppressor genes, Cip1/p21 and p16INK4a, by reducing DNA methylation and increasing histones acetylation in human skin cancer cells. *Carcinogenesis.* 2011;32(4):537-44.
45. Jawaid K, Crane SR, Nowers JL, Lacey M, Whitehead SA. Long-term genistein treatment of MCF-7 cells decreases acetylated histone 3 expression and alters growth responses to mitogens and histone deacetylase inhibitors. *J Steroid Biochem Molecul Biol.* 2010;120(4):164-71.
46. Papoutsis AJ, Lamore SD, Wondrak GT, Selmin OI, Romagnolo DF. Resveratrol prevents epigenetic silencing of BRCA-1 by the aromatic hydrocarbon receptor in human breast cancer cells. *J Nut.* 2010;140(9):1607-14.
47. Shankar S, Srivastava RK. Involvement of Bcl-2 family members, phosphatidylinositol 3'-kinase/AKT and mitochondrial p53 in curcumin (diferulolylmethane)-induced apoptosis in prostate cancer. *Int J Oncol.* 2007;30(4):905-18.
48. Kang S-K, Cha S-H, Jeon H-G. Curcumin-induced histone hypoacetylation enhances caspase-3-dependent glioma cell death and neurogenesis of neural progenitor cells. *Stem Cell Dev.* 2006;15(2):165-74.
49. Shivdasani RA. MicroRNAs: regulators of gene expression and cell differentiation. *Blood.* 2006;108(12):3646-53.
50. Negrini M, Ferracin M, Sabbioni S, Croce CM. MicroRNAs in human cancer: from research to therapy. *J Cell Sci.* 2007;120(11):1833-40.
51. Kim DH, Sætrum P, Snøve O, Rossi JJ. MicroRNA-directed transcriptional gene silencing in mammalian cells. *Proc Natl Acad Sci.* 2008;105(42):16230-5.
52. Benhamed M, Herbig U, Ye T, Dejean A, Bischof O. Senescence is an endogenous trigger for microRNA-directed transcriptional gene silencing in human cells. *Nat Cell Biol.* 2012;14(3):266-75.
53. Marsit CJ, Eddy K, Kelsey KT. MicroRNA responses to cellular stress. *Cancer Res.* 2006;66(22):10843-8.
54. Davis CD, Ross SA. Evidence for dietary regulation of microRNA expression in cancer cells. *Nut Rev.* 2008;66(8):477-82.
55. Pogribny IP, Tryndyak VP, Ross SA, Beland FA. Differential expression of microRNAs during hepatocarcinogenesis induced by methyl deficiency in rats. *Nut Rev.* 2008;66(1):33-5.
56. Tryndyak VP, Ross SA, Beland FA, Pogribny IP. Down-regulation of the microRNAs miR-34a, miR-127, and miR-200b in rat liver during hepatocarcinogenesis induced by a methyl-deficient diet. *Mol Carcinog.* 2009;48(6):479-87.
57. Wang L-L, Zhang Z, Li Q, Yang R, Pei X, Xu Y, et al. Ethanol exposure induces differential microRNA and target gene expression and teratogenic effects which can be suppressed by folic acid supplementation. *Hum Reprod.* 2009;24(3):562-79.
58. Tsang WP, Kwok TT. Epigallocatechin gallate up-regulation of miR-16 and induction of apoptosis in human cancer cells. *J Nut Biochem.* 2010;21(2):140-6.
59. Sun Q, Cong R, Yan H, Gu H, Zeng Y, Liu N, et al. Genistein inhibits growth of human uveal melanoma cells and affects microRNA-27a and target gene expression. *Oncol Rep.* 2009;22(3):563-7.
60. Spurling CC, Suhl JA, Boucher N, Nelson CE, Rosenberg DW, Giardina C. The short chain fatty acid butyrate induces promoter demethylation and reactivation of RAR β 2 in colon cancer cells. *Nutrition and cancer.* 2008;60(5):692-702.

61. Tili E, Michaille J-J, Alder H, Volinia S, Delmas D, Latruffe N, et al. Resveratrol modulates the levels of microRNAs targeting genes encoding tumor-suppressors and effectors of TGF β signaling pathway in SW480 cells. *Biochem Pharmacol*. 2010;80(12):2057-65.
62. Sun M, Estrov Z, Ji Y, Coombes KR, Harris DH, Kurzrock R. Curcumin (diferuloylmethane) alters the expression profiles of microRNAs in human pancreatic cancer cells. *Mol Cancer Therapeut*. 2008;7(3):464-73.
63. Noratto GD, Kim Y, Talcott ST, Mertens-Talcott SU. Flavonol-rich fractions of yaupon holly leaves (*Ilex vomitoria*, Aquifoliaceae) induce microRNA-145 and have anti-inflammatory and chemopreventive effects in intestinal myofibroblast CCD-18Co cells. *Fitoterapia*. 2011;82(4):557-69.
64. Sarveswaran S, Liroff J, Zhou Z, Nikitin AY, Ghosh J. Selenite triggers rapid transcriptional activation of p53, and p53-mediated apoptosis in prostate cancer cells: Implication for the treatment of early-stage prostate cancer. *Int J Oncol*. 2010;36(6):1419-28.